

## REVIEWS

---

# Curative Measures for Cystic Fibrosis: A Perspective on Current Stem Cell-Based, Gene, and Small Molecule Therapies

Priscilla O. Ajilore<sup>1a</sup>, Henry Y. Yang<sup>1</sup>, Anastassia Kerasidis<sup>1</sup>, Ruben Castro<sup>1</sup>

<sup>1</sup> Georgetown University School of Medicine

Keywords: cystic fibrosis, stem cell therapy, gene therapy, small-molecule therapy

<https://doi.org/10.52504/001c.38728>

---

Georgetown Medical Review

Vol. 6, Issue 1, 2022

---

Cystic fibrosis (CF), which is caused by a defect or deficiency in the cystic fibrosis transmembrane conductance regulator (*CFTR*) gene, continues to be a life-limiting multiorgan disease with severe phenotypic manifestations in affected patients. Current approaches to CF therapy have advanced far beyond symptomatic treatment, targeting the aberrant *CFTR* for therapeutic results. Novel small molecule treatments, or CF modulators, were the first to significantly improve the quality of life for patients with CF. These low-molecular-weight drugs can easily traverse the cell membrane and effect transcriptive changes in cells, albeit only for those with the specific mutations addressed by the drugs. However, other stem cell-based treatments, such as mesenchymal stromal cell therapy or induced pluripotent stem cell therapy, and gene therapies, such as CRISPR/Cas9 and viral vectors, are being researched as potential mutation-independent cures. These therapies have yet to progress to clinical trials, but their efficacies in various CF models prove their promise as future treatment options and potential cures. In this review, 3 potential contemporary therapies for CF and their current statuses and trajectories as clinical tools are discussed.

## Introduction

Cystic fibrosis (CF) is a multiorgan disease affecting the exocrine mucous glands of the lungs, liver, pancreas, and intestines.<sup>1</sup> Affecting 1 in every 4000 neonates in the United States, CF continues to pose a significant disease burden.<sup>2</sup> The disease is inherited in an autosomal recessive manner and results from a mutation on chromosome 7 in the cystic fibrosis transmembrane conductance regulator (*CFTR*) gene.<sup>2</sup> Up to 90% of patients with CF have a mutation in the form of a Phe508 deletion, resulting in an amino acid, phenylalanine, being deleted from the CFTR protein. However, numerous other *CFTR* mutations have been identified as causes of CF.<sup>3</sup> The *CFTR* gene encodes an ion channel responsible for transporting chloride and bicarbonate across epithelial cell membranes. Aberrant ion channels along the lumen of respiratory tracts cause significantly reduced ion and water secretion into lumen mucus. This results in excessively thick and stagnant mucus, with a predilection to block the airways and acts as a nidus for infective organisms. Mutations in the lungs are phenotypically expressed as decreased airway clearance, increased airway mucus and subsequent bacterial infection, and lung

---

<sup>a</sup> Corresponding author:  
[poa11@georgetown.edu](mailto:poa11@georgetown.edu)

damage.<sup>1</sup> Although the disease impacts many organs, the respiratory manifestations of the disease are often the focus of therapies because targeting the lungs can significantly improve quality of life and decrease mortality in individuals with CF.<sup>4</sup>

The current management of CF consists of treatments targeted toward symptomatic improvements. However, a curative treatment targeting the underlying pathophysiology of the disease has yet to be clinically realized. This review outlines 3 main curative approaches at the forefront of CF research: stem cell–based therapies, gene therapies, and small molecule treatments. Small molecule treatments have demonstrated sufficient efficacy in human trials.<sup>1</sup> Ninety percent of the disease-causing variants are responsive to current small molecule treatments.<sup>4</sup> Stem cell–based treatments, which include mesenchymal stromal cell (MSC) therapy and induced pluripotent stem cell (iPSC) therapy, as well as gene therapies could potentially be effective in all the disease-causing variants. However, these treatment strategies are still in their infancy, and their safety and feasibility have yet to be determined.

## **Stem Cell Therapies**

### ***Mesenchymal Stromal Cell Therapy***

MSCs are stem cells derived from the bone marrow and found in the umbilical cord and adipose tissue.<sup>5</sup> Due to their dynamic anti-inflammatory properties, MSC therapies could potentially treat lung inflammation in patients with CF.<sup>5</sup> Their anti-inflammatory properties are dynamic in that the environment that the MSCs are exposed to can alter the level of MSC cytokine expression.<sup>6</sup> The ability for MSCs to produce anti-inflammatory signals in response to their environment was demonstrated in a study that involved the introduction of MSCs to bronchoalveolar lavage fluid (BALF) from patients with CF and acute exacerbations or coexisting acute respiratory distress syndrome (ARDS).<sup>6</sup> As a control, patients with CF but without acute exacerbations or coexisting ARDS had their BALF extracted and exposed to MSCs.<sup>6</sup> Notably, the MSCs exposed to patients with CF exacerbations or ARDS had an increase in the production of IL-6, a proinflammatory cytokine that potentially exacerbates neutrophil-induced inflammation and lung damage.<sup>6</sup> In contrast, MSCs exposed to BALF from patients without CF exacerbations or ARDS exhibited significantly reduced IL-6 levels.<sup>6</sup> This understanding of the environmental dynamics of MSCs was an integral step towards conditioning the MSCs as therapeutic agents for CF and other diseases.<sup>6</sup> Thus, it can be inferred that MSCs will produce anti-inflammatory effects and favorable outcomes in patients with CF and lower levels of baseline inflammation.<sup>6</sup>

Unfortunately, the translation of in vitro MSC studies to in vivo applications is not straightforward. Notably, the ability of MSCs to function in the lungs of a patient with CF is compromised when the respiratory environment possesses pathogenic microbial activity.<sup>7</sup> Therefore, researchers recommend that the

lung environments of potential patients are assessed before using MSCs as a therapeutic agent.<sup>7</sup> Being that CF is commonly associated with a large amount of inflammation and mucous secretions in the airways that create an opportune environment for fungi and bacteria, MSC treatment may have decreased efficacy for those with more infectious symptoms.<sup>7</sup> A notable study tested this hypothesis by using similar techniques of introducing MSCs to BALF but instead with patients who had CF and an *Aspergillus* infection.<sup>7</sup> *Aspergillus* produces a toxin known as gliotoxin, which was found to increase MSC death via a mitochondrial damage pathway.<sup>7</sup> This study suggested that while introducing MSCs to patients with CF may be therapeutic, the treatment effect is also limited by the inflammation inductive microorganisms present in the respiratory system, because they can impede the regeneration of lung epithelial cells.<sup>7</sup>

The current research of delivering MSCs as a reliable therapeutic agent for CF remains in consideration for future practices. While MSCs can provide anti-inflammatory effects, the delivery mechanism for this type of treatment for patients with CF is not fully characterized or understood.<sup>5</sup> Furthermore, the change in protein profile under stress-induced or nonstress-induced environments brings both advantages and disadvantages in a clinical setting. Its advantages are that the MSCs can be manipulated to provide a therapeutic effect to patients with CF, but the concern lies in whether this therapeutic effect can be maintained.<sup>6</sup> Additionally, if an infection is present, it may also complicate the anti-inflammatory response by MSCs.<sup>7</sup>

The anti-inflammatory response by MSCs is mediated by the release of their extracellular vesicles, otherwise known as their *secretomes*. To circumvent the challenges of using MSCs entirely, researchers investigated whether extracellular vesicles could be used directly as a potential therapeutic.<sup>8</sup> The extracellular vesicles were found to decrease IL-1B and IL-6 but not IL-8 levels.<sup>8</sup> Thus, researchers concluded that the extracellular vesicles secreted by MSCs were the source of anti-inflammatory effects and could potentially be used directly instead of full MSC transplantation in the lungs.<sup>8</sup> This finding emphasized the constantly changing landscape of MSC research, indicating the recency of the data on treatment and the refinement needed before gaining clinical utility as a curative agent for CF.

### ***Induced Pluripotent Stem Cell Therapy***

The development of iPSC therapy for the treatment of CF holds great potential. iPSCs can be derived from fully differentiated somatic cells, such as skin or blood cells, which are then reprogrammed to an undifferentiated state.<sup>9</sup> Treatment with specific growth factors and signaling molecules can guide subsequent iPSC differentiation into specific epithelial progenitors and ultimately fully differentiated tissue-specific epithelial cells.<sup>9-13</sup>

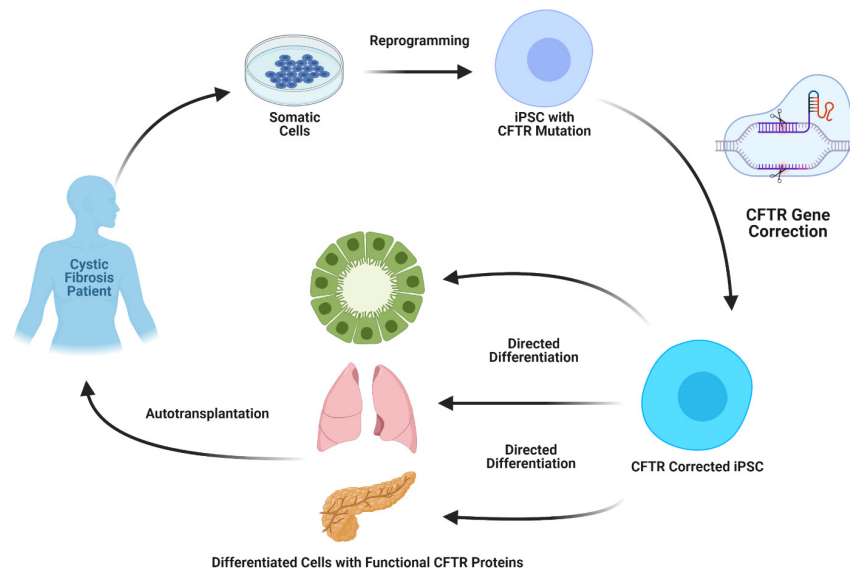
**CFTR Gene Correction in Cystic Fibrosis Induced Pluripotent Stem Cells**

Figure 1. Cystic Fibrosis (CF) Transmembrane Conductance Regulator Gene Correction in Induced Pluripotent Stem Cells

Reprogramming techniques are used to generate induced pluripotent stem cells (iPSCs) from the somatic cells of a patient with CF.<sup>10</sup> Genome-editing technology is used to correct the CF transmembrane conductance regulator (CFTR) mutation responsible for the disease, followed by directed differentiation, yielding fully differentiated cell types with functional CFTR proteins.<sup>14</sup> In theory, the cells would then be transplanted back to the same patient to replace defective tissue. Figure created with [BioRender.com](https://www.biorender.com).

Like human embryonic stem cells (ESCs), iPSCs have the capacity for self-renewal and in vitro differentiation into cell lineages of all 3 embryonic germ layers.<sup>9</sup> Because iPSCs can be generated from somatic cell types, bypassing the need to destroy human embryos, iPSCs provide an ethical alternative to the use of human ESCs. Moreover, because these cells can be patient-specific, the risk of immunogenic transplant rejection and the need for immunosuppression therapy, in theory, could potentially be eliminated, though research on this treatment modality is still in its infancy.

iPSC therapy in the context of CF would involve using somatic cells obtained from an affected individual to generate populations of patient-specific iPSCs.<sup>10</sup> The iPSCs would then be directed to differentiate into tissue-specific progenitor cells in vitro and, after mutation correction using genome-editing technology and further cell differentiation, reintroduced into the donor.<sup>14</sup> In theory, this treatment would essentially replace the diseased or defective tissue with iPSC-derived epithelial cells expressing functional wild-type CFTR proteins, restoring full tissue function ([Figure 1](#)).

The first hurdle in developing pluripotent cell–based therapies to treat CF is developing a replicable and reliable method of generating large populations of progenitor cells capable of differentiation into tissue-specific epithelial cell types. Researchers have demonstrated success in generating patient-specific functional airway epithelial cells from pluripotent stem cells, such as ESCs

and iPSCs, using signaling pathways similar to those involved in in vivo embryologic lung development.<sup>10,11</sup> Using a carefully timed sequence of growth factors and signaling molecules mimicking the stages of embryologic development, Mou et al<sup>10</sup> successfully produced iPSC-derived lung and airway progenitors capable of generating respiratory epithelium when subcutaneously transplanted into immunodeficient mice.

Although the primary focus of iPSC research has been on generating lung epithelium, the potential use of iPSCs for the regeneration of other organ tissues affected by CF is also being investigated. For example, researchers have generated fully differentiated cholangiocytes, epithelial cells lining bile ducts, from iPSCs obtained from patients with CF.<sup>12</sup> Moreover, Simsek and colleagues<sup>13</sup> demonstrated successful differentiation of wild-type and CF iPSCs into pancreatic ductal epithelial cells. Together, these findings suggest that, in addition to lung tissue, iPSC-based therapies may hold potential for the restoration of *CFTR* function in other tissues damaged by CF as well.

The true potential of iPSCs in the treatment and cure of CF centers around their use in combination with gene-editing technology, discussed in later sections of this review, to correct the mutations responsible for the defective *CFTR* protein. Using CRISPR/Cas9 gene-editing systems, researchers have demonstrated successful correction of the F508 *CFTR* gene deletion in CF iPSCs, yielding fully differentiated lung epithelial cells expressing functional *CFTR* proteins.<sup>15</sup> Crane et al<sup>14</sup> were able to correct the same mutation in CF iPSCs using zinc-finger nucleases (ZFNs) with similar results. While these findings are promising, the F508 deletion is one of many mutations responsible for the development of CF. Therefore, more research is needed to assess whether the efficacy of these methods would extend to other genotypes.

While pluripotent stem cell therapies for CF hold potential, many questions regarding their safety and clinical application remain to be answered. In addition to the safety concerns associated with genomic editing, the inherent potential for self-renewal and pluripotency of iPSCs raise concerns about potential tumorigenicity associated with iPSC-based therapies. Furthermore, the process by which gene-corrected iPSC-derived tissue will be transplanted back into the patient remains to be elucidated. Extensive research is needed before iPSC therapies for CF are prepared for clinical trials.

## Gene Therapy

Advances in gene therapy provide promise towards finding a cure for CF. Gene therapy presents the opportunity to correct the underlying genetic error. Unlike *CFTR* modulators, a form of small molecule therapy, gene therapy has the potential to become a curative treatment in all phenotypic presentations of CF.<sup>1</sup> However, there are many barriers to gene delivery that have yet to be overcome to make gene therapy a staple in CF treatment. Namely, pulmonary gene therapy has lagged in clinical development due to the numerous intra- and

extracellular barriers. Although, inhaled gene transfer agents are able to achieve high drug levels in the airways without significant systemic adverse effects, barriers such as the sputum layer coating the airways of patients with CF impede their efficacy.<sup>1,16,17</sup> Fortunately, the advent of drug delivery systems, such as the self-emulsifying drug delivery system, can help gene transfer agents achieve higher mucus permeability.<sup>18</sup> Furthermore, effective treatment is predicated by the delivery mechanism's ability to permanently alter progenitor epithelial cells of the lungs or continually deliver the genes to perpetually replaced terminally differentiated cells.<sup>1,19</sup> Therefore, only a few methods of gene therapy have achieved clinically significant outcomes. The various gene therapy mechanisms are depicted in [Figure 2](#). The research into these contemporary methods is still in its nascent stages, so the potential utility in these mechanisms for curing CF is yet to be fully understood or evaluated.

### ***Viral Gene Transfer Mechanisms***

Given the natural tropism of viruses to the respiratory tract, viruses were extensively studied as promising vectors for *CFTR* gene transfer. To apply this treatment clinically, the viral vector must be capable of efficient gene transfer and sufficient penetration of the enhanced mucus layer in CF fibrosis.<sup>20</sup> However, despite numerous studies with different viruses, no viral vector has demonstrated all the requirements needed for clinical use.<sup>1</sup> The location of receptors on the basolateral surface of the respiratory epithelium and the limitations resulting from preexisting or induced immune responses have resulted in poor viral vector gene transfer efficacy.<sup>1,21</sup>

Many viruses have been investigated as potential vectors for CF gene therapy. Among the first studied were the adenoviral vectors, which were shown to be unsuitable for gene transfer due to the lack of apical coxsackie-adenovirus receptors on human respiratory epithelium.<sup>21</sup> Adeno-associated viral vectors were also investigated, only to show similarly unremarkable results when used to treat the maxillary sinuses of patients with CF due to their limited carrying capacity.<sup>22</sup> Lentiviral vectors have the most promise as a CF treatment because they can transfect dividing and nondividing cells and have demonstrated a longer-term gene expression in their targeted cells.<sup>1</sup> However, they carry the potential risk of insertional mutagenesis, and the research for this vector on CF specifically has yet to be conducted.<sup>1</sup>

In summary, viral vectors have yet to show considerable efficacy to be considered for CF treatment. Although lentiviruses have the most potential as an effective delivery mechanism, their use in CF research has not progressed to clinical trials, underlining the unsuitability of current viral mechanisms for CF gene therapy.

## CURATIVE GENE THERAPIES FOR CYSTIC FIBROSIS

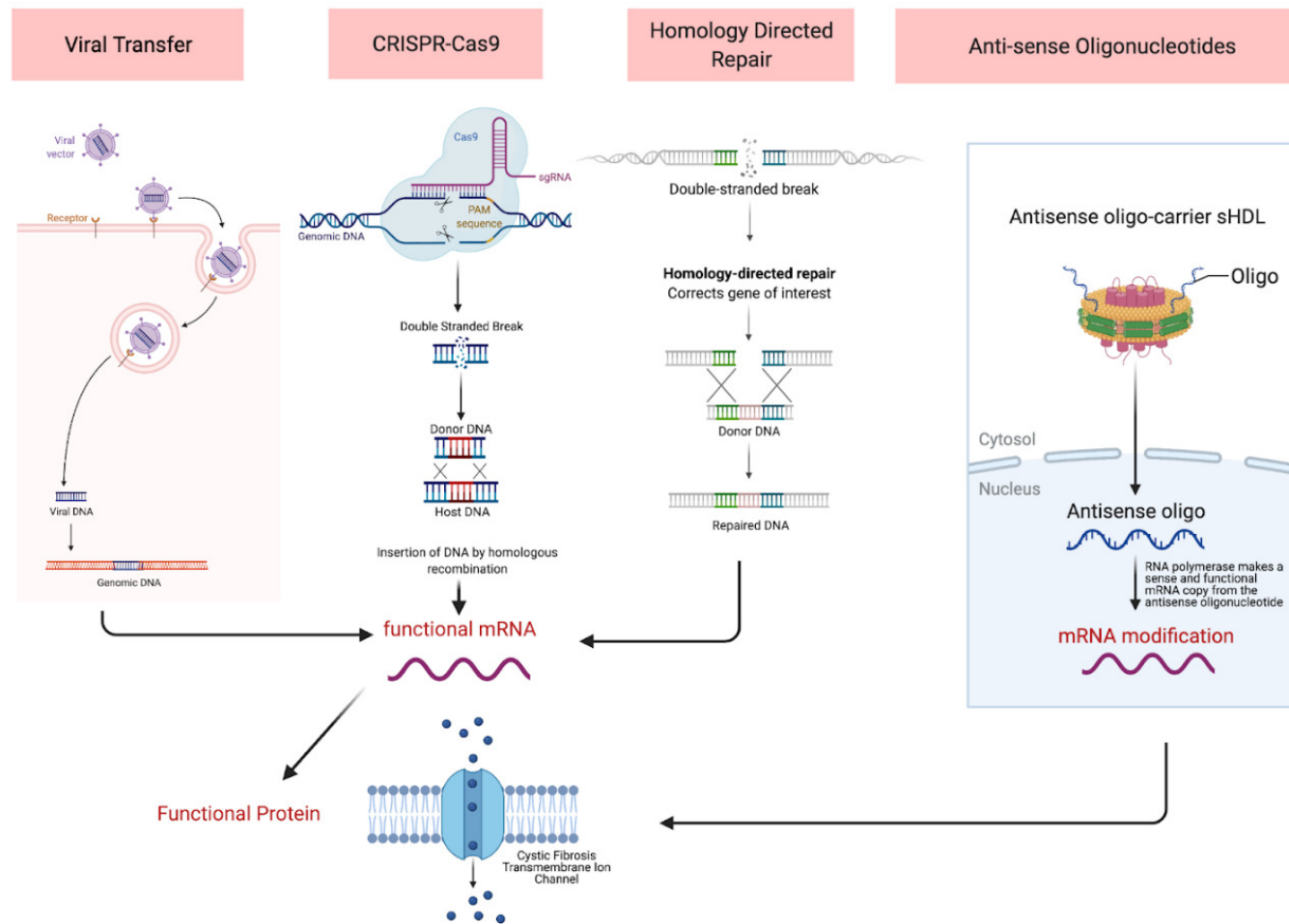


Figure 2. Current Gene Therapies for Cystic Fibrosis.

Gene therapy involves the correction of the CF transmembrane conductance regulator (*CFTR*) defect. There are numerous ways in which the genome of respiratory epithelia can be altered. They include viral vector transfer, CRISPR/Cas9 gene editing, homology-directed repair, and antisense oligonucleotides, which is the only method that modulates the genetic expression on the level of mRNA.<sup>4</sup> The resulting cells after therapy would then express functional *CFTR* proteins, reversing the disease process of CF. Figure created with [BioRender.com](https://www.biorender.com).

## ***Nonviral Gene Transfer Mechanisms***

Unlike their viral counterparts, nonviral vectors are not limited by packaging capacity or immune response.<sup>1</sup> They include naked DNA, cationic liposomes, and DNA nanoparticles or polymers.<sup>1</sup> A phase 1/2a study conducted by Alton et al<sup>23</sup> investigating liposome-mediated gene therapy for patients with CF depicted a significant treatment effect and stabilization of lung function for patients who received the therapy.<sup>1</sup> However, the investigators did not proceed to phase 3 studies due to the demonstrated variability in the treatment effects.<sup>1</sup> Although these vectors have data showing their potential efficacy in CF treatment, they cannot transfect the apical surface of airway epithelial cells efficiently and are limited in their ability to traverse the cytoplasm to reach the nucleus.<sup>1</sup>

## ***Homology-Directed Repair and CRISPR***

Therapies using homology-directed repair (HDR) use the natural HDR process of the cell to correct the defective *CFTR* gene.<sup>24</sup> HDR involves creating a double-stranded break on 2 ends of a target site and replacing the defective gene with a homologous and endogenous wild-type sequence.<sup>24</sup> The most significant caveat of this method is that HDR occurs rarely, only 1 in 10<sup>6</sup> times, making it extremely difficult to experiment with and an ineffective treatment.<sup>24</sup>

Difficulties in the use of the viral and nonviral delivery mechanisms spurred the development of target integration using nucleases to provide a more pointed approach in correcting the defective *CFTR* gene locus. As opposed to therapy using retroviral vectors, programmable nucleases can target DNA sequences at specific cleavage sites.<sup>25</sup> Targeted editing tools, such as ZFN, TALEN, and most commonly CRISPR/Cas9, are being studied for potential use in CF treatment.<sup>24</sup>

Whereas the previous mechanisms involved the delivery of functional genes, gene editing can correct the underlying genetic defect at its original locus. The ability to fix the core defect has the advantages of maintaining the endogenous promoter, allowing for sustained gene expression and natural regulation, and bypassing the risks of insertional mutations associated with foreign DNA.<sup>1</sup> CRISPR/Cas9 is regarded as the best in genomic editing because its efficiency and simplicity far surpass that of ZFNs or transcription activatorlike effector nucleases.<sup>1</sup> Studies have explored the potential of synthesizing stem cells with CRISPR/Cas9 gene editing in curing CF. For instance, Schwank et al<sup>26</sup> found that CRISPR/Cas9 was able to correct the genetic defect at the *CFTR* locus in adult stem cells and provided lasting genetic and functional correction to the resulting clonally expanded organoids. However, the research only suggests the plausibility of inserted CRISPR/Cas9-corrected stem cells into the lungs of patients with CF; in reality, there are no current trials affirming the viability or



functionality of these cells in humans. Although the promise of a permanent fix is tempting, the usage of gene-editing mechanisms as a cure for CF requires much more investigation before it can be tested in human trials.

### ***Antisense Oligonucleotide Therapies***

Messenger RNA (mRNA) is another prime target that can be used in the treatment of CF. Antisense oligonucleotides (ASOs) are used to target gene regions with incorrect splicing instructions, modulating gene expression at the mRNA level, resulting in a functional protein.<sup>2,4</sup> In cystic fibrosis, ASOs can be used to target the splicing mutation 3849 + 10 kb C T. This splice mutation causes the insertion of 84 new nucleotides and a subsequent nonsense mutation that culminates in a truncated CFTR protein.<sup>4</sup> Studies of murine models demonstrated functional protein production after the administration of ASOs.<sup>4,27</sup> ASOs can also be used to insert missing bases in the defective *CFTR* gene locus.<sup>4</sup> Therapy based on mRNA is complicated by its transient nature; *CFTR* restoration through these methods is temporary, and treatment must be administered consistently for the entire length of the patient's lifetime for the potential of consistent results.<sup>2</sup>

### **Small Molecule Therapy**

There have been many pharmacologic interventions and innovations for the treatment of CF, notably *CFTR* modulators, which target the cause of the disease, misfolded CFTR proteins, as opposed to merely treating the symptoms.<sup>28</sup> The first of these drugs to be approved by the Food and Drug Administration (FDA), ivacaftor, demonstrated a considerable improvement in lung function over placebo for patients with the G551D-*CFTR* missense mutation, a class III mutation found in approximately 4% to 6% of patients with CF.<sup>1,29</sup> Ivacaftor allows the ion channel protein to open in the cell membrane.<sup>28</sup> Although this drug was later approved for additional mutations as a monotherapy, ivacaftor's limited therapeutic applicability to the broader CF population demonstrates the current issue with small molecule treatment: a lack of scope.<sup>1</sup> Therefore, although these treatments are currently available for clinical use and have demonstrable treatment efficacies, their usages, especially as monotherapies, are limited to small subsectors of patients with CF with the specific mutations covered by the therapy.<sup>1</sup>

With the numerous possible mutations in the *CFTR* locus, small molecule treatments can be used with one another to maximize efficacy. This additive benefit was shown with the FDA approval of Orkambi (Vertex Pharmaceuticals), a combination of ivacaftor and lumacaftor, in 2015.<sup>1</sup> By adding the *CFTR* corrector lumacaftor, which increases the number of the functional CFTR at the cell surface, the treatable population with small molecule therapy was significantly increased to cover most patients with CF.<sup>1</sup> However, the combination of the 2 has been questioned for their clinical efficacy. The potentially antagonistic drug-drug interaction may explain the only modest improvements in lung function, which were at times eclipsed

by the adverse effects for patients.<sup>1</sup> A newer combination with tezacaftor, a next-generation *CFTR* corrector similar to lumacaftor, was also FDA approved for CF treatment and is expected to have a better benefit-to-risk profile than Orkambi for a larger population of patients with CF.<sup>1</sup> The ivacaftor and tezacaftor combination has not yet had any reports of the same issues faced by Orkambi, showing promise for improved patient treatment outcomes with continued therapy.<sup>1</sup>

Another more recent combination drug, Trikafta (Vertex Pharmaceuticals), is a combination of ivacaftor, tezacaftor, and elexacaftor.<sup>3</sup> All 3 compounds in this combination drug work on various components of the *CFTR* protein to stabilize it. Trikafta is approved for anyone carrying 1 copy of the F508 deletion, even if their second allele holds a different mutation.<sup>3</sup>

Further exploration into other additive therapies is currently underway, with many other *CFTR*-focused drugs currently in clinical trials.<sup>1</sup> New combinations of *CFTR* stabilizers, potentiators, and correctors are currently being tested by pharmaceutical companies for greater efficacy across larger proportions of the CF population and improved safety profiles, including VX-445 and VX-659.<sup>1</sup> Unlike the other categories of therapies previously mentioned, small molecule treatment is the only one with current clinical usage. The greatest challenge to its use, however, remains the applicability to the entire CF population. Outside of the patients with CF who have current FDA-approved treatments, ie, patients with the F508del, there are other patients who do not benefit from small molecule treatments due to the rarity of their mutations.<sup>1</sup>

## Discussion

The management of CF has improved significantly since FDA approval of the first *CFTR* modulator, ivacaftor. Although small molecule treatments are not currently approved for the entire CF population, they were the first to provide treatment for the disease that extended beyond mere symptomatic relief.<sup>1</sup> Through the combination of various *CFTR* modulators, new drugs such as Orkambi and Trikafta represent the current trajectory of the pharmacologic treatment of patients with CF.<sup>3,28</sup> However, these drugs still have modest efficacy and do not have the same curative potential as the stem cell-based or gene therapies presented.<sup>1</sup>

The combination of contemporary stem cell research and gene-editing mechanisms provides hope for an eventual cure for CF, but none of the drugs are currently in a clinically relevant state. MSCs can alter their protein expression behavior with respect to changing environments of human lung tissue, but the anti-inflammatory effects of MSC transplantation may be hindered by patients with CF because of these stress-induced environments.<sup>7</sup> From a clinical perspective, assessing practical candidates may pose an issue due to the dynamicity of a patient's disease state.<sup>7</sup> Further, obtaining BALF

from a patient could be detrimental in relation to time consumption, resource availability, and patient satisfaction. While EVs secreted by the MSCs can avoid the hurdles posed by transplantation, a sustained treatment response with them will still require lifelong treatment.<sup>8</sup> MSCs should still be considered for future CF therapeutic purposes, but new developments and techniques are required to optimize their efficacy.

Further advances in stem cell research have also made it possible to use somatic cells to generate patient-specific self-renewing pluripotent cells capable of directed differentiation into the specific epithelial cell types involved in the pathophysiology of CF.<sup>9-13</sup> When used in combination with genome-editing technology, these techniques can yield new epithelial tissue with fully functional CFTR proteins.<sup>14,15</sup> In theory, this disease-free tissue could then be transplanted back to the donor with minimal risk of transplant rejection. While the development of iPSC-based therapies for the treatment and potential cure of CF is promising, more research is needed to determine the safety and clinical feasibility of these techniques.

## Conclusions

Novel gene-editing mechanisms, such as the use of CRISPR/Cas9 against the progenitor cells of the respiratory tract or novel lentiviral-mediated gene transfer, provide a window to a possible cure for CF. With further clinical trials, these gene therapies have the greatest potential to provide long-lasting CF treatment because of their ability to target the root genetic cause. Therefore, it is conceivable that, once the delivery mechanisms of genomic-editing molecules and the technical and safety barriers behind their usages are determined, they will lead the forefront of CF treatment alongside other stem cell–based methods.

## REFERENCES

1. Schneider-Futschik EK. Beyond cystic fibrosis transmembrane conductance regulator therapy: a perspective on gene therapy and small molecule treatment for cystic fibrosis. *Gene Ther.* 2019;26(9):354-362. doi:10.1038/s41434-019-0092-5
2. Michaels WE, Bridges RJ, Hastings ML. Antisense oligonucleotide-mediated correction of CFTR splicing improves chloride secretion in cystic fibrosis patient-derived bronchial epithelial cells. *Nucleic Acids Res.* 2020;48(13):7454-7467. doi:10.1093/nar/gkaa490
3. Middleton PG, Mall MA, Dřevínek P, et al. Elexacaftor-tezacaftor-ivacaftor for cystic fibrosis with a single phe508del allele. *N Engl J Med.* 2019;381(19):1809-1819. doi:10.1056/nejmoa1908639
4. Maule G, Arosio D, Cereseto A. Gene therapy for cystic fibrosis: progress and challenges of genome editing. *Int J Mol Sci.* 2020;21(11):E3903. doi:10.3390/ijms21113903
5. Caretti A, Peli V, Colombo M, Zulueta A. Lights and shadows in the use of mesenchymal stem cells in lung inflammation, a poorly investigated topic in cystic fibrosis. *Cells.* 2019;9(1):E20. doi:10.3390/cells9010020
6. Abreu SC, Rolandsson Enes S, Dearborn J, et al. Lung inflammatory environments differentially alter mesenchymal stromal cell behavior. *Am J Physiol Lung Cell Mol Physiol.* 2019;317(6):L823-L831. doi:10.1152/ajplung.00263.2019
7. Abreu SC, Hampton TH, Hoffman E, et al. Differential effects of the cystic fibrosis lung inflammatory environment on mesenchymal stromal cells. *Am J Physiol Lung Cell Mol Physiol.* 2020;319(6):L908-L925. doi:10.1152/ajplung.00218.2020
8. Zulueta A, Colombo M, Peli V, et al. Lung mesenchymal stem cells-derived extracellular vesicles attenuate the inflammatory profile of cystic fibrosis epithelial cells. *Cell Signal.* 2018;51:110-118. doi:10.1016/j.cellsig.2018.07.015
9. Takahashi K, Tanabe K, Ohnuki M, et al. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell.* 2007;131(5):861-872. doi:10.1016/j.cell.2007.11.019
10. Mou H, Zhao R, Sherwood R, et al. Generation of multipotent lung and airway progenitors from mouse escs and patient-specific cystic fibrosis iPSCs. *Cell Stem Cell.* 2012;10(4):385-397. doi:10.1016/j.stem.2012.01.018
11. Wong AP, Bear CE, Chin S, et al. Directed differentiation of human pluripotent stem cells into mature airway epithelia expressing functional CFTR protein. *Nat Biotechnol.* 2012;30(9):876-882. doi:10.1038/nbt.2328
12. Ogawa M, Ogawa S, Bear CE, et al. Directed differentiation of cholangiocytes from human pluripotent stem cells. *Nat Biotechnol.* 2015;33(8):853-861. doi:10.1038/nbt.3294
13. Simsek S, Zhou T, Robinson CL, et al. Modeling cystic fibrosis using pluripotent stem cell-derived human pancreatic ductal epithelial cells. *Stem Cells Transl Med.* 2016;5(5):572-579. doi:10.5966/sctm.2015-0276
14. Crane AM, Kramer P, Bui JH, et al. Targeted correction and restored function of the CFTR gene in cystic fibrosis induced pluripotent stem cells. *Stem Cell Reports.* 2015;4(4):569-577. doi:10.1016/j.stemcr.2015.02.005
15. Firth AL, Menon T, Parker GS, et al. Functional gene correction for cystic fibrosis in lung epithelial cells generated from patient iPSCs. *Cell Rep.* 2015;12(9):1385-1390. doi:10.1016/j.celrep.2015.07.062

16. Schuster BS, Kim AJ, Kays JC, et al. Overcoming the cystic fibrosis sputum barrier to leading adeno-associated virus gene therapy vectors. *Mol Ther*. 2014;22(8):1484-1493. [doi:10.1038/mt.2014.89](https://doi.org/10.1038/mt.2014.89)
17. Agent P, Parrott H. Inhaled therapy in cystic fibrosis: agents, devices and regimens. *Breathe*. 2015;11(2):110-118. [doi:10.1183/20734735.021014](https://doi.org/10.1183/20734735.021014)
18. Griesser J, Hetényi G, Federer C, et al. Highly mucus permeating and zeta potential changing self-emulsifying drug delivery systems: a potent gene delivery model for causal treatment of cystic fibrosis. *Int J Pharm*. 2019;557:124-134. [doi:10.1016/j.ijpharm.2018.12.048](https://doi.org/10.1016/j.ijpharm.2018.12.048)
19. Gill DR, Hyde SC. Delivery of genes into the CF airway. *Thorax*. 2014;69(10):962-964. [doi:10.1136/thoraxjnl-2014-205835](https://doi.org/10.1136/thoraxjnl-2014-205835)
20. Mitomo K, Griesenbach U, Inoue M, et al. Toward gene therapy for cystic fibrosis using a lentivirus pseudotyped with Sendai virus envelopes. *Mol Ther*. 2010;18(6):1173-1182. [doi:10.1038/mt.2010.13](https://doi.org/10.1038/mt.2010.13)
21. Walters RW, Grunst T, Bergelson JM, Finberg RW, Welsh MJ, Zabner J. Basolateral localization of fiber receptors limits adenovirus infection from the apical surface of airway epithelia. *J Biol Chem*. 1999;274(15):10219-10226. [doi:10.1074/jbc.274.15.10219](https://doi.org/10.1074/jbc.274.15.10219)
22. Wagner JA, Reynolds T, Moran ML, et al. Efficient and persistent gene transfer of AAV-CFTR in maxillary sinus. *Lancet*. 1998;351(9117):1702-1703. [doi:10.1016/s0140-6736\(05\)77740-0](https://doi.org/10.1016/s0140-6736(05)77740-0)
23. Alton EFWF, Armstrong DK, Ashby D, et al. Repeated nebulisation of non-viral CFTR gene therapy in patients with cystic fibrosis: a randomised, double-blind, placebo-controlled, phase 2b trial. *Lancet Respir Med*. 2015;3(9):684-691.
24. Smirnikhina SA, Kondrateva EV, Adilgereeva EP, et al. P.F508del editing in cells from cystic fibrosis patients. *PLoS ONE*. 2020;15(11):e0242094. [doi:10.1371/journal.pone.0242094](https://doi.org/10.1371/journal.pone.0242094)
25. Jaffé A, Prasad SA, Larcher V, Hart S. Gene therapy for children with cystic fibrosis--who has the right to choose? *J Med Ethics*. 2006;32(6):361-364. [doi:10.1136/jme.2005.01274026](https://doi.org/10.1136/jme.2005.01274026)
26. Schwank G, Koo BK, Sasselli V, et al. Functional repair of CFTR by CRISPR/Cas9 in intestinal stem cell organoids of cystic fibrosis patients. *Cell Stem Cell*. 2013;13(6):653-658. [doi:10.1016/j.stem.2013.11.002](https://doi.org/10.1016/j.stem.2013.11.002)
27. Friedman KJ, Kole J, Cohn JA, Knowles MR, Silverman LM, Kole R. Correction of aberrant splicing of the cystic fibrosis transmembrane conductance regulator (CFTR) gene by antisense oligonucleotides. *J Biol Chem*. 1999;274(51):36193-36199. [doi:10.1074/jbc.274.51.36193](https://doi.org/10.1074/jbc.274.51.36193)
28. Wainwright CE, Elborn JS, Ramsey BW, et al. Lumacaftor-Ivacaftor in patients with cystic fibrosis homozygous for Phe508del CFTR. *N Engl J Med*. 2015;373(3):220-231. [doi:10.1056/nejmoa1409547](https://doi.org/10.1056/nejmoa1409547)
29. Stern M, Ulrich K, Geddes DM, Alton EW. Poly (D, L-lactide-co-glycolide)/DNA microspheres to facilitate prolonged transgene expression in airway epithelium in vitro, ex vivo and in vivo. *Gene Ther*. 2003;10(16):1282-1288. [doi:10.1038/sj.gt.3301994](https://doi.org/10.1038/sj.gt.3301994)